

LUTEOLIN AND THIOSALICYLATE INHIBIT HgCl₂ AND THIMEROSAL-INDUCED VEGF RELEASE FROM HUMAN MAST CELLS

S. ASADI¹, B. ZHANG^{1,2}, Z. WENG¹, A. ANGELIDOU¹, D. KEMPURAJ¹,
K.D. ALYSANDRATOS¹ and T.C. THEOHARIDES^{1,2,3}

¹Molecular Immunopharmacology and Drug Discovery Laboratory, Department of Pharmacology and Experimental Therapeutics, Tufts University School of Medicine and Tufts Medical Center, Boston; ²Department of Biochemistry, Tufts University School of Medicine, Boston; ³Department of Internal Medicine, Tufts University School of Medicine and Tufts Medical Center, Boston, MA, USA

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HgCl₂ is a known environmental neurotoxin, but is also used as preservative in vaccines as thimerosal containing ethyl mercury covalently linked to thiosalicylate. We recently reported that mercury chloride (HgCl₂) can stimulate human mast cells to release vascular endothelial growth factor (VEGF), which is also vasoactive and pro-inflammatory. Here we show that thimerosal induces significant VEGF release from human leukemic cultured LAD2 mast cells (at 1 μM 326±12 pg/10⁶ cells and 335.5±12 pg/10⁶ cells at 10 μM) compared to control cells (242±21 pg/10⁶ cells, n=5, p<0.05); this effect is weaker than that induced by HgCl₂ at 10 μM (448±14 pg/10⁶ cells) (n=3, p<0.05). In view of this finding, we hypothesize that the thiosalicylate component of thimerosal may have an inhibitory effect on VEGF release. Thimerosal (10 μM) added together with the peptide Substance P (SP) at 2 μM, used as a positive control, reduced VEGF release by 90%. Methyl thiosalicylate (1 or 10 μM) added with either SP or HgCl₂ (10 μM) inhibited VEGF release by 100%, while sodium salicylate or ibuprofen had no effect. Pretreatment for 10 min with the flavonoid luteolin (0.1 mM) before HgCl₂ or thimerosal completely blocked their effect. Luteolin and methyl thiosalicylate may be useful in preventing mercury-induced toxicity.

Mercury compounds are found in fish, such as tuna, due to water pollution as well as in antiseptics, disinfectants, toothpastes, lens solutions, and various drugs such as contraceptives, fungicides, and herbicides. Ethyl mercury covalently linked to thiosalicylate, known as thimerosal, has been extensively used as a preservative in vaccines (1). Mercury is a known neurotoxin associated with neurodevelopmental defects and peripheral neuropathies. Mercury exposure causes immune, sensory, neurological, motor, and behavioral

dysfunction similar to symptoms associated with autism (1).

We recently reported that HgCl₂ can induce release of VEGF from human mast cells. HgCl₂ also induces the release of histamine and cytokines, such as IL-4 and tumor necrosis factor-alpha (TNF-α) from a murine mast cell line and mouse bone marrow-derived cultured mast cells (2).

In this study, we investigated whether thimerosal could stimulate VEGF release from human mast cells, and whether its thiosalicylate component and

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Mailing address: T.C. Theoharides, Ph.D., M.D.
Department of Pharmacology and Experimental Therapeutics,
Tufts University School of Medicine,
136 Harrison Avenue, Boston, MA 02111, USA
Tel: ++1 617 636-6866 Fax: ++1 617 636-2456
e-mail: theoharis.theoharides@tufts.edu

the flavonoid luteolin could block this effect.

MATERIALS AND METHODS

Materials

HgCl₂ was obtained from Fluka Chemical Corp. (Milwaukee, WI). Thimerosal, sodium salicylate, methyl thiosalicylate, SP, ibuprofen and luteolin were obtained from Sigma-Aldrich (St. Louis, MO, USA). HgCl₂ was dissolved in Dulbecco's phosphate buffered saline (DPBS, GIBCO, Grand Island, NY, USA). Luteolin was dissolved in DMSO, but the final concentration was less than 0.1%. The other drugs were dissolved in distilled water on the day of the experiments.

Human mast cell culture

LAD2 human mast cells were cultured in serum-free media (StemPro-34; GIBCO, Grand Island, NY, USA) supplemented with 2 mM L-glutamine, 100 IU/ml penicillin, 50 µg/ml streptomycin, and 100 ng/ml recombinant human Stem Cell Factor (rhSCF) (3). For optimal cell growth, LAD2 cell density was maintained between 0.5×10⁶ and 1×10⁶ cells/ml. Cell viability was assessed at 1 h and 24 h using trypan blue (0.3%) exclusion.

VEGF assay

LAD2 cells were washed with DPBS and suspended in complete culture medium. LAD2 cells (2×10⁵ cells/well/200µl) were plated in 96-well flat bottom Falcon cell culture plates (Becton Dickinson, Franklin Lakes, NJ, USA) and were pre-incubated for 15 min at 37°C in 5% CO₂ atmosphere. The cells were then incubated with either SP (2 µM), HgCl₂ (10 µM) or thimerosal (1, 10 µM) for 24 h at 37°C. Control cells were treated with equal volume of only culture medium. For the inhibition experiments, thimerosal or methyl thiosalicylate were added together with the triggers, while luteolin was added 10 min before the triggers. After the reaction time, plates were centrifuged and the supernatant was gently collected from the wells and stored at -20°C until VEGF was measured by Enzyme-Linked Immunosorbent Assay (ELISA) using a commercial kit (R&D Systems, Minneapolis, MN, USA). The minimum detectable level of VEGF was 5 pg/ml.

Statistical analysis

All conditions were performed in triplicate, and all experiments were repeated up to five times (n=3-5). Results are presented as mean±SD. Data from two conditions, such as between stimulated and control samples, were compared using the unpaired 2-tailed, Student's t-test. Significance of comparisons is denoted by p<0.05.

RESULTS

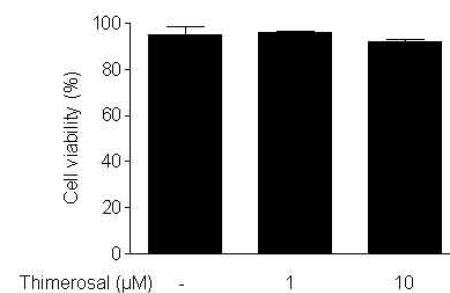
Effect of thimerosal on mast cell viability

LAD2 mast cells were incubated with thimerosal for 24 h in the culture medium and their viability was assessed by trypan blue exclusion. Viability was decreased by less than 10% only at thimerosal concentrations of 10 µM (n=3, Fig. 1A).

Effect of thimerosal on LAD2 mast cell VEGF release

LAD2 cells stimulated with thimerosal for 24 h released significantly more VEGF at 1 and 10

1A



1B

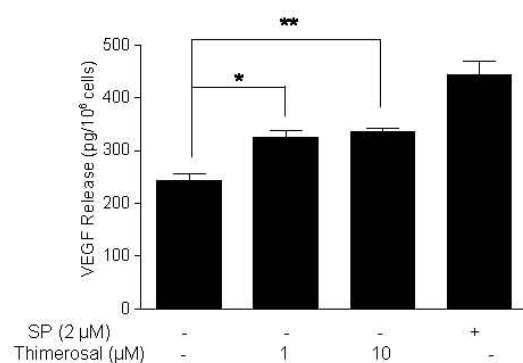
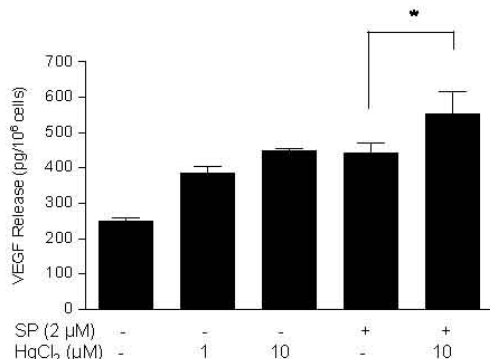


Fig 1. A) The effect of thimerosal on mast cell viability after 24 h. **B)** Effect of thimerosal on VEGF release from LAD2 mast cells. Mast cells were incubated with thimerosal and/or SP as indicated for 24 h for VEGF release at 37°C. The cells were then centrifuged and the supernatant fluid was collected. VEGF release was assayed by ELISA (n=5, p<0.05).

2A



2B

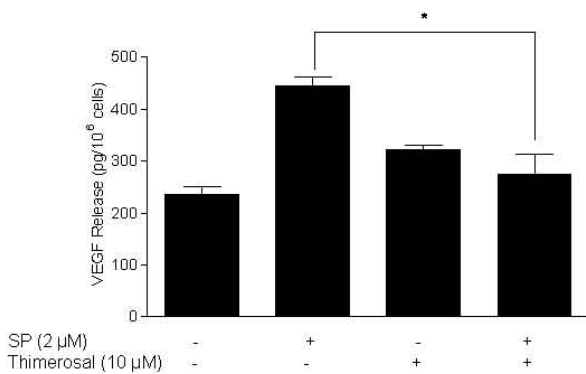


Fig 2. Effect of (A) $HgCl_2$ and (B) thimerosal on SP-induced VEGF release from LAD2 cells. Mast cells were incubated as indicated for 24 h at 37°C. The cells were then centrifuged and the supernatant fluid was collected. VEGF release was assayed by ELISA ($n=3$, $p<0.05$).

μM (326 ± 12 pg/10⁶ cells, 335 ± 12 pg/10⁶ cells), respectively, compared to control (242.5 ± 21 pg/10⁶ cells) ($n=5$, $p<0.05$, Fig. 1B).

Effect of thimerosal on SP-induced VEGF release

$HgCl_2$ (10 μM) added together with SP (2 μM) had a statistically significant effect in augmenting VEGF release (553 ± 63 pg/10⁶ cells) as compared to SP alone (445 ± 16 pg/10⁶ cells) (Fig. 2A). In contrast, thimerosal (10 μM) added together with SP (2 μM) inhibited SP-induced VEGF release to 274.5 ± 35 pg/10⁶ cells ($n=3$, $p<0.05$, Fig. 2B). There was no statistical difference between thimerosal alone and

3A

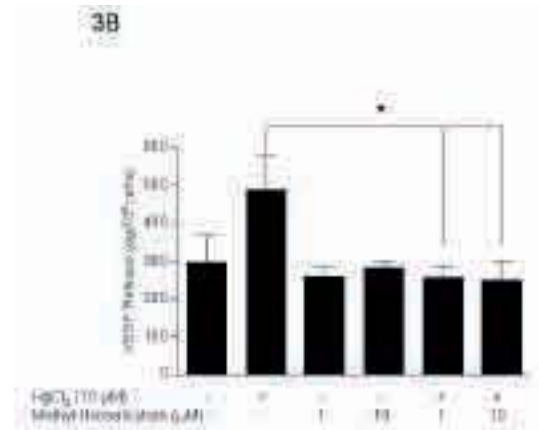
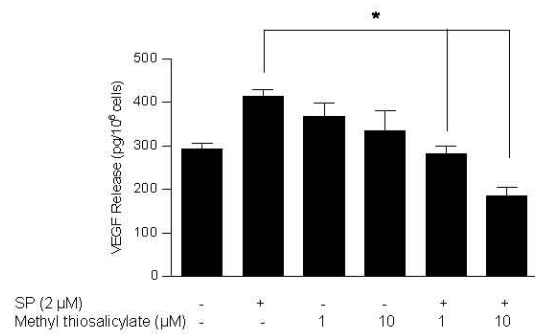


Fig 3. Effect of methyl thiosalicylate on (A) SP and (B) $HgCl_2$ -induced VEGF release. Mast cells were incubated with methyl thiosalicylate and SP or $HgCl_2$ for 24 h at 37°C. The cells were then centrifuged and the supernatant fluid was collected. VEGF release was assayed by ELISA ($n=3$, $p<0.05$).

thimerosal with SP.

Effect of methyl thiosalicylate on $HgCl_2$ and SP-induced VEGF release

In view of the fact that thimerosal inhibited the effect of SP, we hypothesized that the inhibition was due to its thiosalicylate component. We therefore examined the effect of methyl thiosalicylate on SP-induced VEGF release. Methyl thiosalicylate (1, 10 μM) added with the trigger inhibited SP (2 μM)-induced VEGF release from 413.72 ± 18 pg/10⁶ cells to 281.63 ± 24 pg/10⁶ cells at 1 μM and 185.85 ± 23 pg/10⁶ cells at 10 μM (Fig. 3A). Methyl thiosalicylate

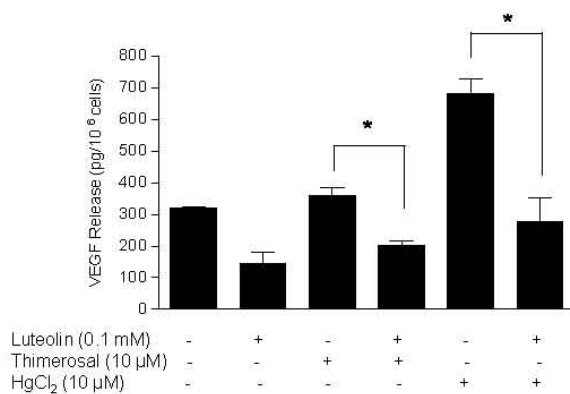


Fig 4. Effect of pretreatment with luteolin on thimerosal and HgCl₂-induced VEGF release. VEGF release was assayed by ELISA (n=3, p<0.05).

(1, 10 μM) also inhibited HgCl₂ (10 μM)-induced VEGF release from 488±76 pg/10⁶ cells to 258±29 and 249.5±24 pg/10⁶ cells, respectively (n=3, p<0.05, Fig. 3B).

Effect of luteolin on thimerosal and HgCl₂-induced VEGF release

Luteolin is a flavonoid known to inhibit mast cell secretion. Therefore, we examined whether luteolin could inhibit thimerosal - or HgCl₂-induced VEGF release. Pretreatment (10 min) of LAD2 cells with luteolin (0.1 mM) blocked VEGF release (100% inhibition) stimulated by either thimerosal (10 μM) or HgCl₂ (10 μM) (n=3, p<0.05, Fig. 4).

DISCUSSION

This is the first report to our knowledge showing that luteolin and methyl thiosalicylate can inhibit SP and HgCl₂-induced VEGF release from human cultured mast cells. The mechanism of thiosalicylate inhibition is not clear. Sodium salicylate and ibuprofen had no effect at the same concentration as thiosalicylate, implying that cyclooxygenase is not involved. Luteolin and thiosalicylate may work by preventing intracellular calcium elevations since thimerosal increased cytosolic calcium levels in thymus lymphocytes (4) as HgCl₂ did in PC12 cells (2).

Mast cells are potential targets for environmental agents with immunotoxic effects because they

are mostly located in the skin, respiratory and gastrointestinal tracts (5). Mast cells are critical for allergic reactions, for innate and acquired immunity (6), as well as in inflammation (7). Non-allergic mast cell triggers can derive from either the gut or the brain (7), and include neuropeptides such as SP (8). Once activated, mast cells secrete numerous vasoactive, pro-inflammatory and neurotoxic molecules; these include histamine, prostaglandin D₂, IL-6 (7) and VEGF (9-10), an isoform of which is vasodilatory and is overexpressed in delayed hypersensitivity reactions (11). In fact, mast cells can release VEGF (12), IL-6 (13) and other mediators “selectively” without degranulation (14), leading to “allergic-like” symptoms not evidenced by typical diagnostic tests. Such mediators could disrupt the blood-brain barrier (BBB) (15-16) permitting brain inflammation and further increasing brain HgCl₂ levels, especially since HgCl₂ can cross the BBB through a transport mechanism (1, 17).

Methyl mercury ingestion from fish was previously linked to neurological damage (18). Exposure to mercury at critical developmental periods (1, 19) may contribute to the pathogenesis of neurodevelopmental disorders such as autism, especially in subjects with autism susceptibility genes (19). Such subjects may be further vulnerable because of mast cell sensitivity or activation by other triggers (20). For instance, a preliminary report indicated that the incidence of autism is 10-fold higher in mastocytosis patients (1/10 children) (21), characterized by increased number of hyperactive mast cells in the skin and other tissues (22), than the general population (1/100 children) (23).

The ability of the natural flavone luteolin (24) to inhibit the effect of SP and HgCl₂ on VEGF release is impressive. Luteolin can inhibit mast cell activation and mast cell-dependent stimulation of activated T cells (25). Luteolin is anti-inflammatory (26), inhibits IL-6 release from microglia (27), and can inhibit autism-like behavior in mice (28). Luteolin (5, 7, 3', 4'-tetrahydroxyflavone) is closely related to 7, 8-dihydroxyflavone recently shown to mimic brain-derived neurotrophic factor (BDNF), which is neuroprotective (29). Luteolin could, therefore, be useful in treating neuroinflammatory diseases either alone or as an adjuvant to other therapeutic approaches (30).

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Disclosure: The authors declare that they have no competing interests. TCT is the inventor of patent application US 12/534,571 and provisional application US61/405,414 covering the use of luteolin in the treatment of autism.

REFERENCES

1. Geier DA, King PG, Sykes LK, Geier MR. A comprehensive review of mercury-provoked autism. *Indian J Med Res* 2008; 128:383-411.
2. Dastyk J, Walczak-Drzewiecka A, Wyczolkowska J, Metcalfe DD. Murine mast cells exposed to mercuric chloride release granule-associated N-acetyl-beta-D-hexosaminidase and secrete IL-4 and TNF-alpha. *J Allergy Clin Immunol* 1999; 103:1108-14.
3. Kirshenbaum AS, Akin C, Wu Y, Rottem M, Goff JP, Beaven MA, Rao VK, Metcalfe DD. Characterization of novel stem cell factor responsive human mast cell lines LAD 1 and 2 established from a patient with mast cell sarcoma/leukemia; activation following aggregation of FcepsilonRI or FcgammaRI. *Leuk Res* 2003; 27:677-82.
4. Elferink JG. Thimerosal: a versatile sulfhydryl reagent, calcium mobilizer, and cell function-modulating agent. *Gen Pharmacol* 1999; 33:1-6.
5. Paus R, Theoharides TC, Arck PC. Neuroimmunoendocrine circuitry of the 'brain-skin connection'. *Trends Immunol* 2006; 27:32-9.
6. Galli SJ, Tsai M. Mast cells in allergy and infection: versatile effector and regulatory cells in innate and adaptive immunity. *Eur J Immunol* 2010; 40:1843-51.
7. Theoharides TC, Kalogeromitros D. The critical role of mast cell in allergy and inflammation. *Ann NY Acad Sci* 2006; 1088:78-99.
8. Theoharides TC, Zhang B, Kempuraj D, et al. IL-33 augments substance P-induced VEGF secretion from human mast cells and is increased in psoriatic skin. *Proc Natl Acad Sci USA* 2010; 107:4448-53.
10. Boesiger J, Tsai M, Maurer M, Yamaguchi M, Brown LF, Claffey KP, Dvorak HF, Galli SJ. Mast cells can secrete vascular permeability factor/vascular endothelial cell growth factor and exhibit enhanced release after immunoglobulin E-dependent upregulation of Fcε receptor I expression. *J Exp Med* 1998; 188:1135-45.
11. Grutzkau A, Kruger-Krasagakes S, Baumeister H, Schwarz C, Kogel H, Welker P, Lippert U, Henz BM, Moller A. Synthesis, storage and release of vascular endothelial growth factor/vascular permeability factor (VEGF/VPF) by human mast cells: Implications for the biological significance of VEGF₂₀₆. *Mol Biol Cell* 1998; 9:875-84.
12. Brown LF, Olbricht SM, Berse B, Jackman RW, Matsueda G, Tognazzi KA, Manseau EJ, Dvorak HF, Van de Water L. Overexpression of vascular permeability factor (VPF/VEGF) and its endothelial cell receptors in delayed hypersensitivity skin reactions. *J Immunol* 1995; 154:2801-7.
13. Cao J, Papadopoulou N, Kempuraj D, Boucher WS, Sugimoto K, Cetrulo CL, Theoharides TC. Human mast cells express corticotropin-releasing hormone (CRH) receptors and CRH leads to selective secretion of vascular endothelial growth factor. *J Immunol* 2005; 174:7665-75.
14. Kandere-Grzybowska K, Letourneau R, Kempuraj D, Donelan J, Poplawski S, Boucher W, Athanassiou A, Theoharides TC. IL-1 induces vesicular secretion of IL-6 without degranulation from human mast cells. *J Immunol* 2003; 171:4830-6.
15. Theoharides TC, Kempuraj D, Tagen M, Conti P, Kalogeromitros D. Differential release of mast cell mediators and the pathogenesis of inflammation. *Immunol Rev* 2007; 217:65-78.
16. Esposito P, Gheorghe D, Kandere K, Pang X, Conally R, Jacobson S, Theoharides TC. Acute stress increases permeability of the blood-brain-barrier through activation of brain mast cells. *Brain Res* 2001; 888:117-27.

17. Esposito P, Chandler N, Kandere-Grzybowska K, Basu S, Jacobson S, Connolly R, Tutor D, Theoharides TC. Corticotropin-releasing hormone (CRH) and brain mast cells regulate blood-brain-barrier permeability induced by acute stress. *J Pharmacol Exp Ther* 2002; 303:1061-6.
18. Burbacher TM, Shen DD, Liberato N, Grant KS, Cernichiari E, Clarkson T. Comparison of blood and brain mercury levels in infant monkeys exposed to methylmercury or vaccines containing thimerosal. *Environ Health Perspect* 2005; 113:1015-21.
19. Harris HH, Pickering IJ, George GN. The chemical form of mercury in fish. *Science* 2003; 301:1203.
20. Deth R, Muratore C, Benzecry J, Power-Charnitsky VA, Waly M. How environmental and genetic factors combine to cause autism: A redox/methylation hypothesis. *Neuro Toxicol* 2008; 29:190-201.
21. Theoharides TC, Kempuraj D, Redwood L. Autism: an emerging 'neuroimmune disorder' in search of therapy. *Expert Opin Pharmacother* 2009; 10:2127-43.
22. Theoharides TC. Autism spectrum disorders and mastocytosis. *Int J Immunopathol Pharmacol* 2009; 22:859-65.
23. Castells M. Mast cell mediators in allergic inflammation and mastocytosis. *Immunol Allergy Clin. North Am* 2006; 26:465-85.
24. Kogan MD, Blumberg SJ, Schieve LA, et al. Prevalence of parent-reported diagnosis of autism spectrum disorder among children in the US, 2007. *Pediatrics* 2009; 5:1395-403.
25. Middleton E Jr, Kandaswami C, Theoharides TC. The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease and cancer. *Pharmacol Rev* 2000; 52:673-751.
26. Seelinger G, Merfort I, Schempp CM. Anti-oxidant, anti-inflammatory and anti-allergic activities of luteolin. *Planta Med.* 2008; 74:1667-77.
27. Jang S, Kelley KW, Johnson RW. Luteolin reduces IL-6 production in microglia by inhibiting JNK phosphorylation and activation of AP-1. *Proc Natl Acad Sci USA* 2008; 105:7534-9.
28. Parker-Athill E, Luo D, Bailey A, Giunta B, Tian, J, Shytle RD, Murphy T, Legradi G, Tan J. Flavonoids, a prenatal prophylaxis via targeting JAK2/STAT3 signaling to oppose IL-6/MIA associated autism. *J Neuroimmunol* 2009; 217:20-27.
29. Jang SW, Liu X, Yepes M, et al. A selective TrkB agonist with potent neurotrophic activities by 7,8-dihydroxyflavone. *Proc Natl Acad Sci USA* 2010; 107:2687-92.
30. Theoharides TC. Luteolin as a therapeutic option for multiple sclerosis. *J Neuroinflammation* 2009; 6:29.